

Full length Research Article

## Cognitive and Neuroprotective Effects of *Vernonia amygdalina* in Scopolamine-induced Memory impaired Rats

\*Odu P.O.<sup>1</sup>, Odu V.K.<sup>1</sup>, Oyebanjo O.T.<sup>2</sup>, Benneth B.<sup>3</sup>, and Onasanwo S.A.<sup>4</sup>

<sup>1</sup>Department of Physiology, Faculty of Basic Medical Sciences, Calabar, Calabar, Nigeria

<sup>2</sup>Department of Physiology, Faculty of Basic Medical Sciences, University of Ibadan, Nigeria

<sup>3</sup>Department of Physiology, Babcock University, Nigeria

<sup>4</sup>Department of Pharmacology and Therapeutics, Faculty of Basic Medical Sciences, College of Health Sciences, Delta State University, Abraka, Delta State, Nigeria.

**Summary:** Cognitive impairment is largely associated with functional and structural loss in the brain of Alzheimer's disease (AD) models, and scopolamine has been successfully used to mimic these deficits in rodents. The cost and side effects of drugs presently used for the treatment of AD-related cognitive impairment have prompted research into alternative products, especially natural ones with high antioxidant capacity since oxidative stress is a major pathophysiology associated with AD. The current study evaluated the cognitive and neuroprotective effects of *Vernonia amygdalina* (VA) on scopolamine-induced cognitive impairment in rats. Thirty-five male rats were randomly divided into seven groups (n = 5). Group 1 served as the control and received distilled water. Groups 2 and 3 received *Vernonia amygdalina*, VA (50 and 100 mg/kg, respectively) per orally (p.o.). Group 4 received 1 mg/kg scopolamine SC (i.p.). Groups 5, 6, and 7 received pretreatment with either VA 50 mg/kg, VA 100 mg/kg, or Donepezil, DP (1 mg/kg), and then in combination with SC (1 mg/kg). The animals were subjected to memory tasks using the Morris water maze (MWM) and novel object recognition tasks (NORT). They were sacrificed on day 14, after which their brains were isolated for biochemical and histological studies. The study showed that during MWM and NORT, spatial and non-spatial recognition memories, respectively impaired in the SC group compared to the control group, were reversed in the VA pretreatment groups. Scopolamine injection caused significant decreases in superoxide dismutase and catalase levels and an increase in malonaldehyde (MDA) levels in group 4 compared with the control group. Pretreatments with either VA or DP, however, caused a significant increase in the SOD and catalase levels and a decrease in the MDA level compared with the SC group. Histological studies revealed that VA was more potent in protecting the brain against SC-induced neurodegeneration and morphological alterations in the hippocampus and prefrontal cortex. The findings of this study suggest that VA attenuates scopolamine-induced cognitive deficits via inhibition of oxidative stress and neuronal degeneration and enhancement of cognition in the brains of rats.

**Keywords:** Cognitive deficits, Alzheimer disease, *Vernonia amygdalina*, Scopolamine, Memory, Oxidative stress.

\*Authors for correspondence: : [odupeterori@unical.edu.ng](mailto:odupeterori@unical.edu.ng), [odupeterori@gmail.com](mailto:odupeterori@gmail.com), Tel: +234-8036865949

Manuscript received- December 2023; Accepted: September 2024

DOI: <https://doi.org/10.54548/njps.v39i2.9>

© 2024 Physiological Society of Nigeria

This article has been published under the terms of Creative Commons Attribution-Non-commercial 4.0 International License (CC BY-NC 4.0), which permits non-commercial unrestricted use, distribution, and reproduction in any medium, provided that the following statement is provided. "This article has been published in the Nigerian Journal of Physiological Sciences.

### INTRODUCTION

Cognitive functions are multiple mental attributes that enable humans and associated organisms to effectively interact with their environment, carry out daily tasks, and adapt to different survival strategies. Amongst these are learning and memory which predominantly have been reported to show a decline in efficiency as a person ages, and in disease conditions like Alzheimer's disease (AD). AD is an age-related, progressive neurodegenerative disorder of the central nervous system, the most common form of dementia characterized by cognitive dysfunction, altered personality, loss of executive function, and ultimately death (Xingxuan *et al.*, 2008; Merlo *et al.*, 2010). Investigations of the brains of AD patients show low levels of neuronal antioxidant defense enzymes, elevated levels of

oxidative stress biomarkers, and increased levels of reactive oxygen species (ROS) and nitrogen species (Walton *et al.*, 2012). Preclinical and clinical studies have revealed extensive neuronal loss and synaptic changes in the basal forebrain, particularly the areas involved in learning and memory (Giacobini, 2003). Scopolamine, an M<sub>1</sub> muscarinic receptor subtype has been experimentally used to induce AD-related disorders in animals. Tota *et al.* (2012) reported that scopolamine impairs memory through mechanisms that increase acetylcholinesterase and induce oxidative stress in the brain. Drugs like donepezil and tacrine currently used to manage AD, are associated with short half-lives, poor bioavailability, and serious side effects, including hepatotoxicity (Rountree *et al.*, 2009). Thus, there is growing research interest in the use of alternative remedies, such as plants with medicinal, antioxidant, and therapeutic

potential, for the treatment of AD-related cognitive deficits. Dietary polyphenols have gained significant attention due to their rich antioxidant and therapeutic potential in reducing somatic cell and neuronal damage (Engelhart *et al.*, 2002). *Vernonia amygdalina* (VA) leaves, commonly known as bitter leaf, have been widely used for both therapeutic and nutritional purposes (Ojiako and Nwanjo, 2006). VA possesses strong polyphenolic, antioxidant, anxiolytic, antinociceptive, and anti-inflammatory (Iwalokun *et al.*, 2006; Onasanwo *et al.*, 2017) properties. It is an abundant source of linoleic and linolenic acids, and polyunsaturated fatty acids found to be protective against cardiovascular diseases (Tapsell, 2006). This study was therefore designed to investigate the cognitive and neuroprotective effects of VA leaf extracts in scopolamine-induced cognitive impairment in Wistar rats.

## MATERIALS AND METHODS

**Experimental animals:** Male Wistar rats (80-120 g) obtained from the central animal house of the University of Ibadan, were used for this study. The rats were housed in plastic cages at room temperature and given free access to commercial food pellets and water *ad libitum*. They were acclimatized for two weeks before commencement of experiments. The experimental procedures were carried out in accordance with the guidelines for animal research as detailed in the guidelines for the Care and Use of Laboratory Animals by the National Research Council of the National Academy of Sciences, 2011.

**Plant and chemicals:** *Vernonia amygdalina*, Scopolamine hydrobromide (Sigma-Aldrich.co., St. Louis, U.S.A), and Donepezil hydrochloride (torrent pharma, UK Ltd.) were used in the study.

**Plant Extraction using Soxhlet technique:** Fresh mature leaves of VA were obtained and verified at the University of Ibadan. The leaves were air-dried and blended into powder form using an electric blender. Its weight (2600 g) was determined using an electric weighing balance and then soaked in 80% (v/v) methanol for 72 hours at a temperature of 40°C. The extract was filtered using a cheese material, and thereafter with Whatman No 1 filter paper. The filtrate was concentrated to dryness using a rotary evaporator.

**Experimental design and treatments:** The rats were divided into 7 groups (n = 5). Group 1 received distilled water (per orally, *p.o*) alone throughout the experiment, from day 1-14 and served as the Control (CT). Group 2 and 3 received *Vernonia amygdalina*, VA (50 and 100 mg/kg respectively) alone (*p.o*) for 14 days. Group 4 received distilled water (*p.o*) from day 1-7 as well as 1 mg/kg scopolamine SC (*i.p*) from day 8-14. Groups 5, 6 and 7 were pretreated with *Vernonia amygdalina* (SC+VA 50 mg/kg), (SC+VA 100 mg/kg), and Donepezil (SC+DP 1 mg/kg) respectively, *p.o* for 7 days, and then in combination with scopolamine (1 mg/kg) (*i.p*) one hour later from day 8-14.

### Behavioral assessments

**Novel object recognition task (NORT):** On the experimental day, animals underwent two sessions, each

consisting of a single trial. In the initial session, referred to as the acquisition trial (trial 1), animals were placed in an arena with two identical objects for 5 minutes. Any rat that did not engage with the objects for at least 20 seconds within the 5-minute period were excluded from the experiments. Exploration was defined as the animal bringing its nose within 1 cm of the object while looking at, sniffing, or touching it. In the second session, known as the retrieval session (trial 2), which occurred 24 hours after trial 1, one of the objects used in the first trial was replaced by a new, unfamiliar object (the novel object). Animals were once again placed in the arena for 5 minutes, during which their total exploration time for each were recorded and the time spent actively exploring the familiar (F) or novel (N) object in trial 2. Olfactory cues were eliminated in between trials by cleaning the test box with 70% of ethanol. Percentage exploratory preference which served as index non-spatial recognition memory was evaluated using the formula:  $[(N)/(N+F)] \times 100$  (Bevins and Besheer, 2006).

**Morris water maze (MWM):** The MWM apparatus used in this study was made of a circular GP water tank (150 cm diameter and 74 cm height) filled with water ( $25 \pm 2$  °C) to a depth of 30 cm. The pool was divided into four hypothetical quadrants, designated as: East, West, North, and South. Throughout the trial, a white round platform of diameter 10 cm was placed 2 cm below the surface of the water in a constant position; in the middle of the southwest quadrant. The water was made opaque by adding a food coloring (peak milk) in order to prevent the animal from noting the depth of the water. Animals were placed 1 inch above the water, positioned to face the wall of the pool. In each trial rats were given a maximum of 60 s cut-off time to locate the hidden platform. This was recorded using a stopwatch, and then averaged for each training session. In the event that the animal was unable to locate the hidden platform within 60 s, it was gently guided to it, and allowed to stay there for 30 s. Each rat was subjected to a daily session of 2 trials for 3 consecutive days, while the animals were allowed to rest on the 4<sup>th</sup> day. On day the 5<sup>th</sup> day, the platform height was further lowered below the water level, and the rats underwent a spatial *probing* trial in which they were given 60 s to search for the platform. Escape latency time (ELT) which was the time taken to locate the hidden platform in the water maze was used as an index of spatial learning and memory (Morris, 1984; Ishola *et al.*, 2013).

**Preparation of Brain Homogenate:** On the 14<sup>th</sup> day of the protocol schedule, animals underwent cervical dislocation. Brains were removed and rinsed with ice-cold isotonic saline solution. Brain tissue samples were then homogenized with 10 times (w/v) ice-cold 0.1M phosphate buffer (pH 7.4). The homogenate was centrifuged at 10,000 rpm for 15 min to obtain the supernatant. The supernatant was collected and separated into different portions that were kept refrigerated for the different biochemical assays.

### Biochemical estimations

**Estimation of Protein concentration:** The protein concentration of brain homogenate was determined by the Biuret method described by Gornal *et al.*, (1949).

**Estimation of Malondialdehyde (MDA):** MDA, an end product of lipid peroxidation was measured by the method described by Varshney and Kale (1990), and expressed in units/mg protein.

**Estimation of Superoxide Dismutase (SOD) Activity:** SOD activity was estimated by the method described by Misra and Frodovich (1972) based on the inhibition of Epinephrine-Adrenochrome transition by the enzyme. The enzyme activity was expressed in terms of units/mg protein.

**Estimation of Nitrite Level:** Nitrite level which served as an indicator of nitric oxide (NO) production was estimated using the Greiss reaction as described by Green *et al.* (1982).

**Histological Studies:** Some of the animals were perfused with 0.01 M phosphate buffer solution (PBS, pH 7.4) followed by 4% paraformaldehyde (PFA) in PBS. Brains were removed and post-fixed in 4% formaldehyde for histological study. The tissues were observed and cut into small pieces not more than 4mm thick into pre-labeled cassettes. The brains were removed and placed in a new formaldehyde solution for 24 h to be fixed before being dehydrated using alcohol (70% for 24 h 80% for 1 h and 95% for 1 h absolute 1 and absolute 11, for 1 hr) then cleared in xylene and embedded in paraffin. Sections were cut with a microtome (Leica TP1020, Germany) at 5 μm thicknesses, mounted on glass slides, and stained with the routine haematoxylin and eosin (H&E) technique stain, and Cresyl fast violet (Nissl). Staining was done as described by Avwioro (2010). Examination of slides, photography, and morphometric studies were done at the Histopathology Department, University Teaching Hospital, Ibadan. Good sections from each group were signed and documented as photomicrographs using Olympus digital camera.

**Statistical Analysis:** All the data were expressed as mean ± standard error of mean, (n=5). Data were analyzed using one- or two-way ANOVA followed by Turkey post hoc test (where applicable) to determine the level of significance. Analysis was done using GraphPad prism software, version 7 (Diego, CA, USA).

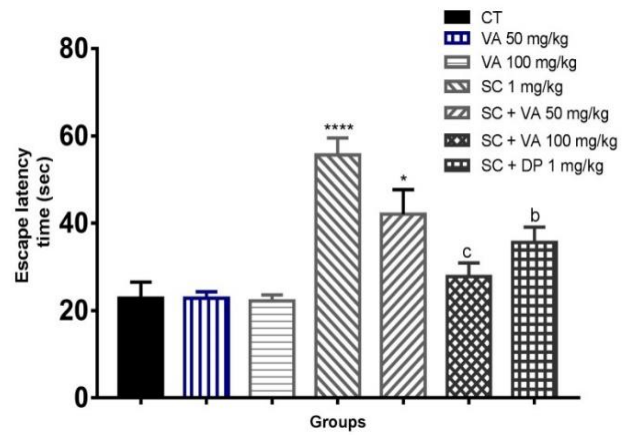
**RESULTS**

**Comparison of percentage exploratory preference (PEP) during NORT in rats:** One-way ANOVA showed significant differences in the PEP across the groups. Post hoc analysis showed that PEP significantly decreased in the SC group (p<0.0001) compared with the control group. However, VA inhibited the effects of scopolamine-induction in the pretreatment groups during NORT test (Fig. 1).

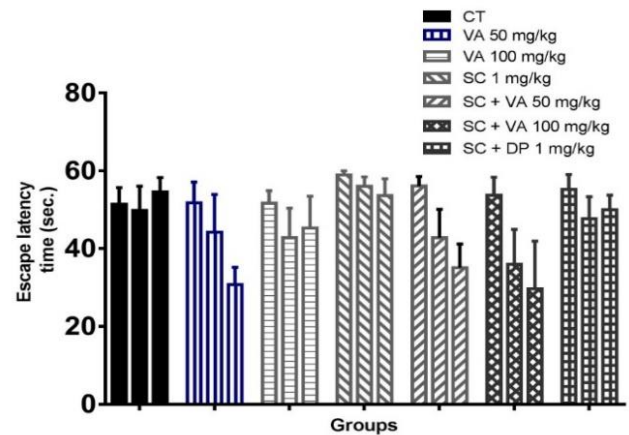
**Comparison of escape latency time (ELT) during acquisition trials in MWM:** One-way ANOVA showed that there were no significant differences in the ELT across the groups during MWM acquisition trials of day 10, 11, and 12. Post hoc analysis showed progressive decreases in ELT in all the groups, except in the group that received scopolamine injection alone (Fig. 2)

**Comparison of ELT during probe trial in MWM:** One-way ANOVA showed significant differences in the ELT to access spatial memory across the groups. Post hoc analysis

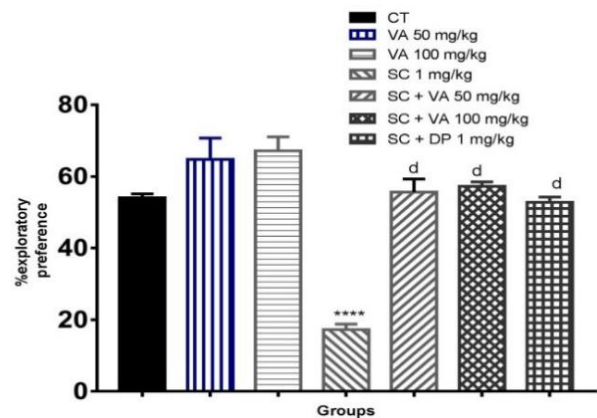
revealed that ELT was significantly increased in the SC group which received scopolamine injection alone (p<0.0001) and in the SC+VA 50 mg/kg (p<0.01) compared with the control group. ELT was however significantly decreased SC+VA 50 mg/kg (p<0.001) and SC+DP 1 mg/kg (p<0.01) compared with the SC group (Fig. 3).



**Figure 1.** PEP during NORT. Values are expressed as mean ± SEM, n = 5. \*\*\*\*p<0.0001 versus Control (CT); <sup>d</sup>p<0.0001 versus scopolamine (SC) treated.

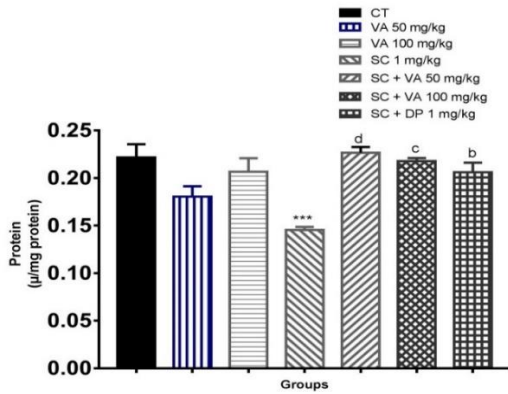


**Figure 2.** Escape latency during acquisition trial in MWM. Values are expressed as Mean ± SEM, n = 5.



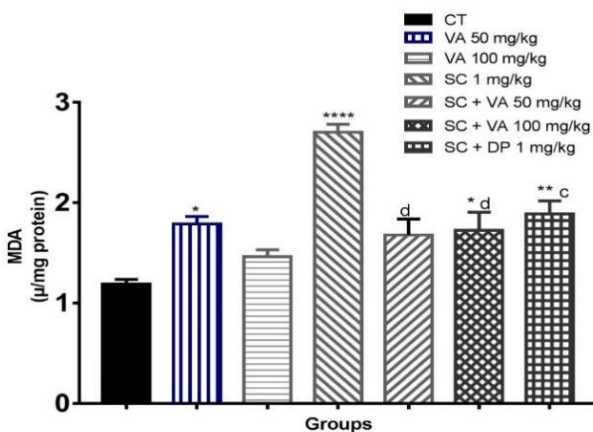
**Figure 3** Escape latency during probe trial in MWM. Values are expressed as mean ± SEM, n = 5. \*p<0.05 and \*\*\*\*p<0.0001 vs CT; <sup>b</sup>p<0.01 and <sup>c</sup>p<0.001 vs SC treated.

**Comparison of protein concentration in experimental rats:** As shown in Fig. 4, one-way ANOVA showed that scopolamine injection produced significant changes in protein concentration. Post hoc analysis showed that Protein concentration decreased significantly ( $p < 0.001$ ) in the SC group compared with the control. However, the SC+VA (50 and 100 mg/kg) and SC+DP pretreatments groups showed significant decreases ( $p < 0.0001$  and  $p < 0.00$ ) and ( $p < 0.01$ ) respectively compared with the SC group (fig. 4).



**Figure 4.** Effects of VA on protein concentration. Values are expressed as mean  $\pm$  SEM,  $n = 5$ . \*\*\* $p < 0.001$  vs CT; <sup>b</sup> $p < 0.01$ , <sup>c</sup> $p < 0.001$  and <sup>d</sup> $p < 0.0001$  vs SC treated.

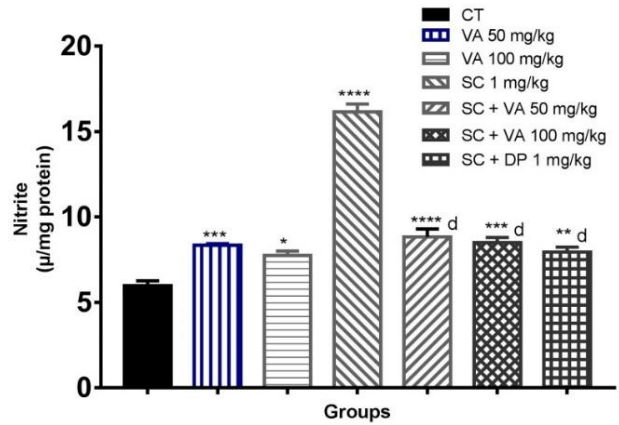
**Comparison of Malonaldehyde (MDA) levels in experimental rats:** Fig. 5 shows the effects of scopolamine injection caused significant changes in the MDA levels in rats' brains as revealed by one-way ANOVA. Post hoc analysis showed that MDA levels significantly increased in the SC ( $p < 0.0001$ ), SC+VA 100 mg/kg ( $p < 0.05$ ), and SC+DP ( $p < 0.001$ ) groups compared with the CT group. While MDA significantly decreased in the SC+VA (50 mg/kg) compared with the SC group, it also decreased significantly in SC+VA 100 mg/kg ( $p < 0.0001$ ), and SC+DP ( $p < 0.001$ ) groups compared with the SC group (fig. 5).



**Figure 5.** Effects of VA on MDA level. \* $p < 0.05$  \*\* $p < 0.01$  and \*\*\*\* $p < 0.0001$  vs CT; <sup>c</sup> $p < 0.001$  and <sup>d</sup> $p < 0.0001$  vs SC treated.

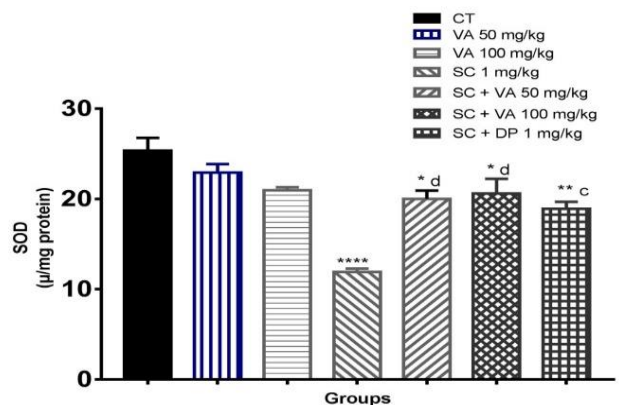
**Comparison of Nitrite levels in experimental rats**  
One-way ANOVA showed significant differences in nitrite levels across the groups. Post hoc analysis showed that nitrite level was significantly increased in SC+VA (50 and 100 mg/kg) groups ( $p < 0.001$ ,  $p < 0.05$ , respectively), and SC

group ( $p < 0.0001$ ) groups compared with the CT group. Nitrite levels also increased significantly in the SC+VA (50 and 100 mg/kg) or SC+DP groups ( $p < 0.0001$ ,  $p < 0.001$ ,  $p < 0.01$ , respectively) compared with the control. However, pretreatment of rats with VA inhibited the severity of scopolamine-induced effects by significant decreases ( $p < 0.0001$ ) in the Nitrite levels across the combination groups (Fig. 6).

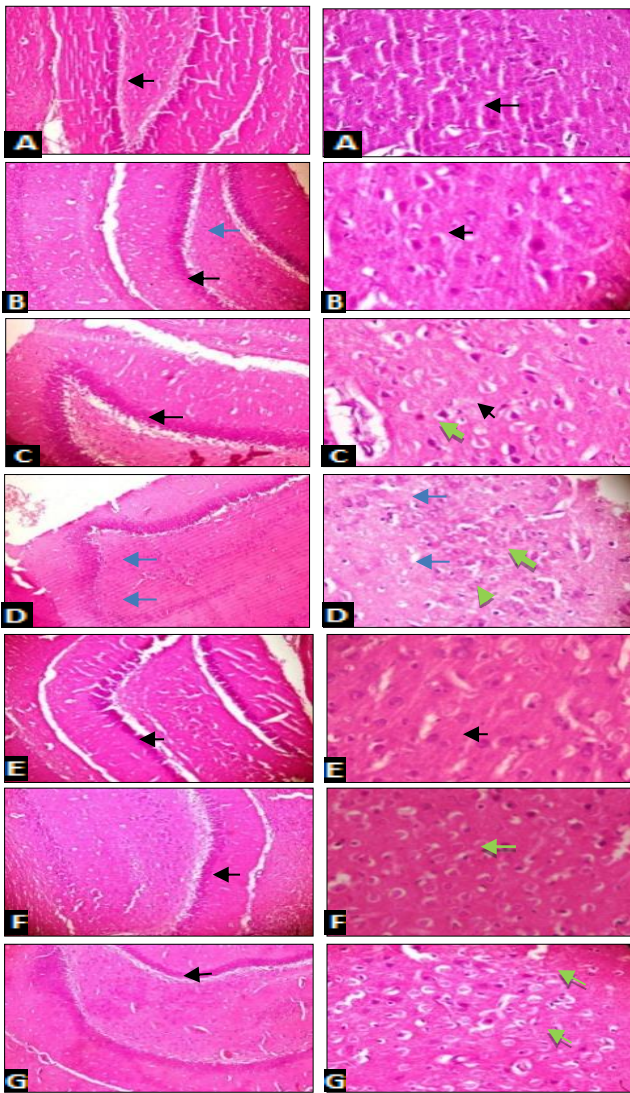


**Figure 6** Effects of VA on Nitrite level. Values are expressed as mean  $\pm$  SEM,  $n = 5$ . \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ , \*\*\*\* $p < 0.0001$  vs CT; <sup>d</sup> $p < 0.0001$  vs SC treated.

**Comparison of SOD level in experimental rats:** One-way ANOVA showed significant differences in the SOD level across the groups. Post hoc analysis showed significant decreases in the SOD levels in the SC group ( $p < 0.0001$ ) compared with the CT group. SOD also showed significant decreases in the SC+VA (50 and 100 mg/kg) and SC+DP groups ( $p < 0.05$  and  $p < 0.05$ ; and  $p < 0.01$ ), respectively compared to the CT group. SOD was however significantly increased SC+VA (50 and 100 mg/kg) and SC+DP groups ( $p < 0.0001$ ); and  $p < 0.001$ ), respectively compared to SC group (Fig. 7).



**Figure 7** Effects of VA on SOD level. \* $p < 0.05$ , \*\* $p < 0.01$ , and \*\*\*\* $p < 0.0001$  vs CT; <sup>c</sup> $p < 0.001$ , and <sup>d</sup> $p < 0.0001$  vs SC treated.

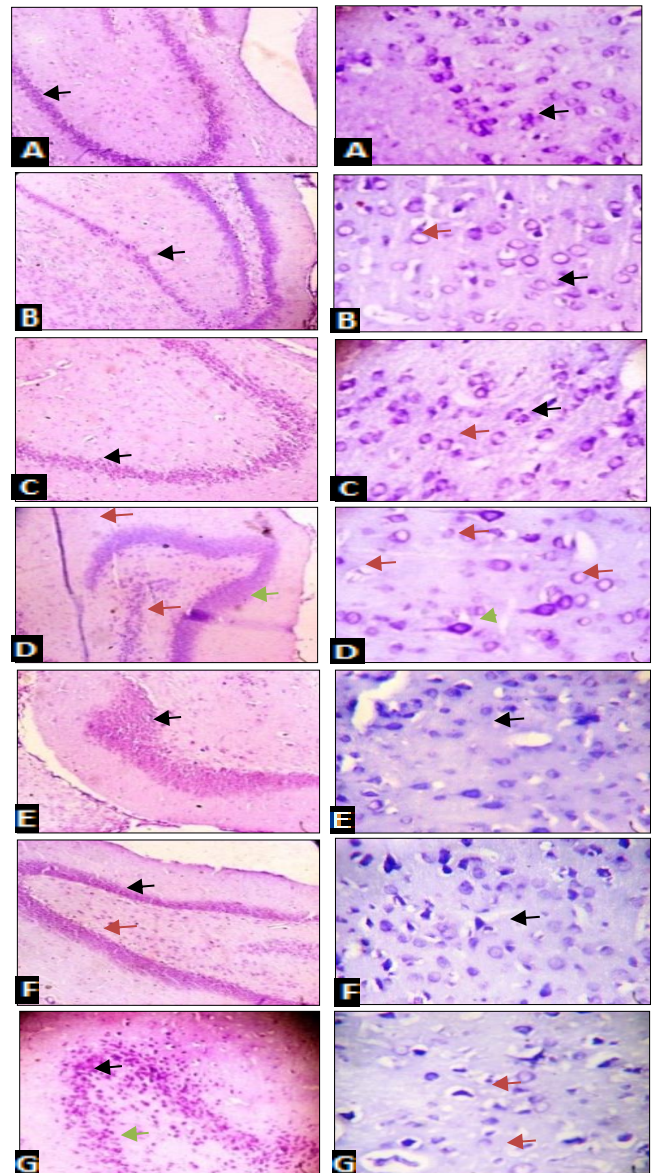


**Plate 1a:** Photomicrograph of brain sections of the **hippocampus, hp** (left), and prefrontal cortex, **pc** (right) stained with H&E.

The Plate 1a shows (A) Control rats exhibit a normal histological structure with intact neuronal cells in both the hippocampus (hp) and prefrontal cortex (pc); (B) Rats treated with *Vernonia amygdalina* (VA, 50 mg/kg) display a few degenerated neurons (hp), and normal neuronal cells (pc); (C) Rats treated with VA (100 mg/kg) show normal neuronal cells (hp), and chromatolysis in a few neuronal cells (pc); (D) scopolamine-induced AD rats showing several degenerated neurons (blue arrows) in both **hp and pc**. Degenerated neurons in the pc appear transparent and exhibit chromatolysis; (E) scopolamine-induced AD rats pretreated with VA treatment (50 mg/kg) showing normal neuronal cells (in hp and pc); (F) scopolamine-induced AD rats pretreated with VA treatment (100 mg/kg) display normal neuronal cells (hp), **and** moderate chromatolysis (pc); (G) scopolamine-induced AD rats pretreated with Donepezil (1 mg/kg) exhibit moderately normal neuronal cells (hp), with some degenerated neuronal cells showing degree of hyalinization and chromatolysis (pc).

Plate 1b shows (A) show a normal distribution of Nissl bodies in neurons within the hippocampus (hp) and prefrontal cortex (pc); (B) VA-treated rats (50 mg/kg) show a moderately normal distribution of Nissl bodies (hp), with

a few neuronal cells displaying loss of Nissl bodies (pc); (C) VA-treated rats (100 mg/kg) display moderately normal distribution Nissl bodies (hp), and some neuronal cells with a loss of Nissl bodies in the pc; (D) Scopolamine-induced AD rats exhibit a poor distribution of Nissl bodies (hp), and significant loss of Nissl bodies (red arrows) with a reduced neuronal population (pc). (E) scopolamine-induced AD rats pretreated with VA (50 mg/kg) show moderately restored distribution of Nissl bodies in both hp and pc. (F) Scopolamine-induced AD rats pretreated with VA (100 mg/kg) show a moderate distribution of Nissl bodies, although a few neurons exhibit a loss of Nissl bodies (hp), some neuronal cells in the prefrontal cortex, particularly within the hypocellular layer, display a loss of Nissl bodies (pc); (G) Scopolamine-induced AD rats pretreated with Donepezil (1 mg/kg) exhibit a moderate distribution of Nissl bodies followed by a low neuronal population (hp), there is a reduction in neuronal cells and a significant loss of Nissl bodies (pc).



**Plate 1b:** Photomicrograph of brain sections of the **hippocampus, hp** (left), and prefrontal cortex, **pc** (right) stained with cresyl fast violet.

## DISCUSSION

Our study showed that scopolamine, a non-selective muscarinic receptor antagonist caused significant damages in neuronal architecture in both the prefrontal cortex and the hippocampus with associated cognitive impairment that relates to AD. Scopolamine alters neuronal processes and facilitates neurodegeneration of cortical and hippocampal neurons. Previous studies have reported that scopolamine interferes with cognitive processes involved in memory consolidation and retrievals thus causing short- and long-term memory impairment (Ishola *et al.*, 2013; Souza *et al.*, 2013). Behavioural, neurochemical, and histological changes induced by scopolamine in the current study were attenuated by pretreatment with *Vernonia amygdalina*.

Scopolamine significantly impaired rodents' recognition memory in the novel object recognition task (NORT), a task widely used to assess rodent's long-term, non-spatial recognition memory (Ennaceur, 2010; Pezze *et al.*, 2017). The NORT takes advantage of rodent's unprompted nature to explore its natural surroundings especially novel objects (Ennaceur, 2010). While a higher time spent with the novel object compared to a familiar one within the rodent's environment indicates good residual memory for the familiar object, the opposite indicates memory deficits. Consequently, pre-treatment with *Vernonia amygdalina* (VA) efficiently prevented memory impairment in the NORT when recognition memory was assessed.

Morris water maze (MWM), a widely used neurobehavioral task, was used to assess spatial memory which is a hippocampal-dependent task. Animals were trained to acquire information about a spatial location and reach the hidden platform in the circular pool. Escape latency time (ELT) which is the time taken to reach the hidden platform was significantly increased following scopolamine treatment indicating impairment in memory. Pretreatment with *Vernonia amygdalina* however inhibited and reversed the severe memory deficits recorded in groups that received only scopolamine.

It has also been reported that the central nervous system is very susceptible to oxidative stress (Walton *et al.*, 2012). Biochemical assays in numerous studies relating to AD shows low levels of neuronal antioxidant defense enzymes and elevated levels of oxidative stress biomarkers, which contributes greatly to neuronal damage and cognitive dysfunction (Walton *et al.*, 2012). In a similar context, scopolamine has been reported in several studies to cause memory impairment via oxidative stress pathology in rodents (Khalifa, 2004; Fan *et al.*, 2005).

The current study also shows that scopolamine-induced oxidative stress significantly causes increases in the brain lipid peroxidation with parallel decreases in total protein concentrations (TPC) and superoxide dismutase activity (SOD). However, pretreatment with VA protected the brain from this oxidative stress impact of scopolamine by causing a decrease in Malonaldehyde (MDA) level, an index of lipid peroxidation MDA level, and parallel increases in TPC and SOD. SOD is an anti-oxidant that detoxifies superoxide anions by converting it to hydrogen peroxide and water (Abhinav, 2010). Although previous studies have shown that the central nervous system is very susceptible to oxidative stress (Walton *et al.*, 2012), the VA was able to exert neuroprotective effects. The rich antioxidant

properties of VA have been reported in previous studies (Owolabi *et al.*, 2008). Ogunlade *et al.* (2012) also reported that SOD and MDA levels significantly improved following *Vernonia amygdalina* treatment in liver oxidative stress caused by alcohol ingestion.

Scopolamine also caused significant elevation in brain Nitric oxide (NO) levels, but this was ameliorated by VA pretreatment. NO is an important oxidative biological signalling molecule with diverse physiological functions including neurotransmission and learning and memory processes (Susswein *et al.*, 2004), however, when generated in excess during oxidative outburst and inflammation, In excess, NO combines with superoxide anion ( $O_2^-$ ) forming the highly reactive and neurotoxic product peroxynitrite (ONOO<sup>-</sup>), which leads to further oxidative and nitrosative stress in part via mitochondrial injury (Rao and Balachandran, 2002; Lipton, 2004). Very high level of NO have been implicated in oxidative stress and neuroinflammation in brain parenchyma, neurodegeneration, and neuronal cell death through NO-mediated neurotoxicity (Law *et al.*, 2004; Togo, 2004).

Histological findings in this study support earlier reports that scopolamine extensively destroys neuronal architecture and hippocampal circuits that may play vital roles in learning and memory (Haroutunian *et al.*, 1997; Choi *et al.*, 2012; Chen *et al.*, 2014), as well as depopulates neuronal cells via neurotoxicity (Ramirez-Rodriguez *et al.*, 2011). It has also been reported that scopolamine attenuates memory task-induced increases of regional cerebral blood flow in the prefrontal cortex and the right anterior cingulate region (Grasby *et al.*, 1995). The histological report of the current study showed that scopolamine caused neurodegeneration, chromatolysis, hyalination several losses of nissl bodies in the hippocampus and prefrontal cortex.

Pretreatment with VA however protected the brain from the severity of scopolamine-induced damages. While VA was able to reduce the level of damage induced by scopolamine exposure, there were however other negative side effects across the groups with VA treatment. This may be due to the presence of bioactive compounds such as oxalate previously reported to be insoluble in the methanol form of VA extractions (Nerurkar *et al.*, 2004; Eleyinmi *et al.*, 2006). The presence of oxalate in food has been associated with acidity and toxicity (Eleyinmi *et al.*, 2006). However, studies that involved an aqueous form of *Vernonia amygdalina* extraction recorded no histopathological conditions (Ogunlade *et al.*, 2012; Mebratu *et al.*, 2013). On the other hand, the donepezil treatment group was associated with adverse negative side effects characterized by undifferentiated layers, degeneration, chromatolysis, reduction in neuronal cells, and loss of Nissl bodies, thus making it less effective to protect the brain compared with VA. Moreover, a study by Zaki *et al.* (2013) reported that donepezil was not strong enough in protecting the brain against scopolamine insults and was associated with some levels of degeneration (Zaki *et al.*, 2013). Donepezil works via a cholinergic pathway to inhibit scopolamine activity and enhance acetylcholine levels which improves cognition. While the scope of this study didn't consider this pathway, VA may be working via the same route to execute its beneficial function.

In conclusion, VA demonstrated neuroprotective effects and improved cognitive deficits induced by scopolamine in

rats thus suggesting a beneficial role in conditions associated with cognitive dysfunctions. Although the cognitive enhancing effect of VA was parallel to that of donepezil, the underlying pathological adverse effects reported in the present and previous studies limit its wide usage. Further research into the mechanism via which VA executes its function may position it as a better alternative to donepezil.

## REFERENCES

- Abhinav, K., Jogender, M., Madhusudana, K., Vegi, G., Modi, N., Yogendra, K. and Gupta, R. (2010). Anti-Amnesic Activity of Vitex Negundo in Scopolamine Induced Amnesia in Rats. *Pharmacology & Pharmacy*, 1, 1-8.
- Allam, A.R., Kiran, K., and Hanuman, T. (2008). Bioinformatic analysis of Alzheimer's Diseases using functional protein sequences. *J Proteomics Bioinform*, 1, 036-042.
- Ballard, C.G., Chalmers, K.A., Todd, C., McKeith, I.G., O'Brien, J.T., Wilcock, G., Love, S. and Perry, E.K. (2007). Cholinesterase inhibitors reduce 673 cortical Abeta in dementia with Lewy bodies. *Neurology*, 674 68:1726-1729.
- Bevins, R.A. and Besheer, J. (2006). Object recognition in rats and mice: a one-trial non-matching-to-sample learning task to study 'recognition memory'. *Nature Protocls*.1(3):1306-11.
- Bruin N. and Pouzet, B. (2006). Beneficial effects of galantamine on performance in the object recognition task in Swiss mice: Deficits induced by scopolamine. *Pharmacology, Biochemistry and Behavior* 85: 253-260.
- Chen, L., Richardson, J., Caldwell, J. and Ang, L. (1994). Regional brain activity of free radical defense enzymes in autopsy samples from patients with Alzheimer's disease and from non-demented controls, *International Journal of Neuroscience* 75, 83-90.
- Choi, D.Y., Lee, Y.J., Lee, S.Y., Lee, Y.M., Lee, H.H., and Choi, I.S. (2012). Attenuation of scopolamine-induced cognitive dysfunction by obovatol. *Arch Pharm Res.*, 35:1279-86.
- Easton, A., Sankaranarayanan, S., Tanghe, A., Terwel, D., Lin, A. X., Hoque, N., Bourin, C., Gu, H., Ahlijanian, M. and Bristow, L. (2013). Effects of sub- 700 chronic donepezil on brain Abeta and cognition in a mouse model 701 of Alzheimer's disease. *Psychopharmacology*, 230:279-289.
- Eleyinmi, A., Adebowale, Y., Oluwalana, I., Ajisafe, O. and Akintomide, T. (2006). Effect of dietary inclusion of Garcinia kola, Gongronema latifolium and Vernonia amygdalina on the nutritional quality of a complementary diet. *Journal of Biological Sciences.*, 1:43-49.
- Engelhart, M.J., Geerlings, M.I., Ruitenber, A., van Swieten J.C., Hofman, A., Witteman, J.C., et al. (2002). Dietary intake of antioxidants and risk of Alzheimer disease. *JAMA*. 287:3223-9
- Ennaceur, A. (2010). One-trial object recognition in rats and mice: methodological and theoretical issues. *Behav. Brain Res.*, 215(2), 244-54.
- Fan Y, Hu J, Li J, Yang Z, Xin X, Wang J, et al. (2005). Effect of acidic oligosaccharide sugar chain on scopolamine-induced memory impairment in rats and its related mechanisms. *Neurosci Lett*; 374(3):222-6.
- Grasby, P.M., Frith, C.D., Paulesu, E., Friston, K.J., Frackowiak, R.S. and Dolan, R.J. (1995). The effect of the muscarinic antagonist scopolamine on regional cerebral blood flow during the performance of a memory task. *Exp Brain Res.*, 104:337-348.
- Giacobini, E. (2003). Cholinesterases: new roles in brain function and in Alzheimer's disease. *Neurochemical Research*, 28, 515- 522.
- Gornall, A. G., Barddawill, C. J. and David, M. M. (1949). Determination of serum proteins by means of biuret reaction. *J Bio Chem.*, 177(2): 751-66.
- Green, L.C., Wagner, D.A., Glogowski, J., Skipper, P.L., Wishnok, J.S. and Tannenbaum, S.R. (1982). Analysis of nitrate, nitrite, and [15N] nitrate in biological fluids. *Anal Biochem* 126(1):131-8
- Guo, H. B., Cheng, Y. F., Wu, J. G., Wang, C. M., Wang, H. T., Zhang, C., Qiu, Z. K. and Xu, J. P. (2015). Donepezil improves learning and memory deficits in app/ps1 mice by inhibition of microglial activation. *Neuroscience* 30, 20-30.
- Haroutunian, V., Greig, N., Pei, X.F., Utsuki, T., Gluck, R. and Acevedo, L.D. (1997). Pharmacological modulation of Alzheimer's bamyloid precursor protein levels in the CSF of rats with forebrain cholinergic system lesions. *Brain Res Mol Brain Res*, 46:161-8
- Herbet, V. (1996). Prooxidant effects of antioxidant vitamins. *Introduction J. Nutr.*126 (4 Suppl): 1197S-200S.
- Ishola, I. O., Adamson, F. M. and Adeyemi O. O. (2016). Ameliorative effect of kolaviron, a biflavonoid complex from Garcinia kola seeds against scopolamine-induced memory impairment in rats: role of antioxidant defense system. *Metab Brain Dis*
- Ishola, I. O., Tota, S., Adeyemi, O. O., Agbaje, E. O., Narender, T. and Shukla, R. (2013). Protective effect of Cnestis ferruginea and its active constituent on scopolamine-induced memory impairment in mice: a behavioral and biochemical study. *Pharm Biol.*, 51(7):825-835.
- Iwalokun, B. A., Efedede, B. U., Alabi-Sofunde, J. A., Odualu, T., Magbagveola, O. A. and Akinwande, A. I. (2006). Hepatoprotective and antioxidant activities of Vernonia amygdalina on acetaminophen-induced hepatic damage in mice. *J. Med. Food.*, 9(4): 524-530.
- Katzman, R., and Saitoh, T. (1991). Advances in Alzheimer's disease. *FASEB J* 5: 278-286.
- Kaur, R., Parveen, S., Mehan, S., Khanna, D. and Kalra, S. (2015). Neuroprotective effect of Ellagic acid against chronically scopolamine induced Alzheimer's Type Memory and Cognitive Dysfunctions: Possible Behavioural and Biochemical Evidences *International Journal of Preventive Medicine Research*. (1): 2, 45-64.
- Khalifa AE. (2004). Pro-oxidant activity of zuclopenthixol in vivo differential effect of the drug on brain oxidative status ofscopolamine-treated rats. *Hum Exp Toxicol*; 23 (9):439-45.
- Kumar, A., Dogra, S. and Prakash, A. (2009). Neuroprotective Effects of Centella asiatica against Intracerebroventricular Colchicine Induced Cognitive Impairment and Oxidative Stress. *Int J Alzheimers Dis.*, 9, (7): 21-78.
- Law, A., Gauthier, S. and Quirion, R. (2004). Say NO to Alzheimer's disease: The putative links between nitric oxide and dementia of the Alzheimer's type. *Brain Res Brain Res Rev.*, 35: 73-96
- Lipton, S.A. (2004). Concepts: turning down but not off – neuroprotection requires paradigm shift in drug development. *Nature*, 428, 473.
- Mebratu, A., Yamrot, K., Eyasu, M., Yonas, B., and Kelbesa, U. (2013). Toxic effects of aqueous leaf extracts of Vernonia bipontini Vatke on blood, liver and kidney tissues of mice. *Monona Ethiopian Journal of Science (MEJS)*, 5:15-31.
- Merlo, s., Spampinato, S., Canonico, P. L., Copani, A. and Sortino M.A. (2010). Alzheimer's disease: brain expression

- of a metabolic disorder? Trends in Endocrinology and Metabolism 21: 537–544.
- Misra, H. P. and Fridovich, I. (1972). The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. *J Biol Chem.*, 25; 247(10):3170-5.
- Morris, R. G. (1984). "Development of a water maze procedure for studying spatial learning in the rat." *J Neurosci Methods.*, 11:47–60.
- Nerurkar, P., Dragull, K. and Tang, C. (2004). In vitro toxicity of kava alkaloid, pipermethystime, in Hep G2 cells compared to kavalactones. *Journal of Toxicological Sciences*, 79:106-111.
- Ogunlade, B., Akunna, G. G., Fatoba, O. O., Ayeni, O. J., Adegoke, A. and Adelokun, A. (2012). *Vernonia amygdalina* Protects Against Hepatotoxicity in Wistar Rats. *World J Young Researchers*, 2(5)2(5):71
- Ojiako, O.A. and Nwanjo, H.U. (2006). Is *Vernonia amygdalina* hepatotoxic or hepatoprotective? Response from biochemical and toxicity studies in rats. *Africa Journal of Biotechnology*, 5(18): 1648-1651.
- Onasanwo, S.A., Oyebanjo, O.T., Ajayi, A.M. and Olubori M.A. (2017). Anti-nociceptive and anti-inflammatory potentials of *Vernonia amygdalina* leaf extract via reductions of leucocyte migration and lipid peroxidation (2017). *J Intercult Ethnopharmacol.* 6:2.
- Owolabi, M.A., Jaja, S.I., Oyekanmi, O.O. and Olatunji O.J. (2008). Evaluation of the antioxidant activity and lipid peroxidation of the leaves of *Vernonia amygdalina*. *J. Compl. Integr. Med.*, 5.
- Rao, A.V. and Balachandran, B. (2002). Role of Oxidative Stress and Antioxidants in Neurodegenerative Diseases. *Nutritional Neuroscience*, 5(5):291-309.
- Pezze, M., Marshall, H. J. and Cassaday H. J. (2017.) Scopolamine Impairs Appetitive But Not Aversive Trace Conditioning: Role of the Medial Prefrontal Cortex. *The Journal of Neuroscience*, 37(26):6289 – 6298.
- Ramirez-Rodriguez, G., Ortiz-Lopez, L., Dominguez-Alonso, A., Benitez-King, G.A. and Kempermann, G. (2011). Chronic treatment with melatonin stimulates dendrite maturation and complexity in adult hippocampal neurogenesis of mice. *J Pineal Res* 50:29-37.
- Rountree, S. D., Chan, W, Pavlik, V. N., Darby, E. J., Siddiqui, S. and Doody, R. S. (2009). Persistent treatment with cholinesterase inhibitors and/or memantine slows clinical progression of Alzheimer disease. *Alzheimers Res Ther.*, 1:7.
- Souza, A.C., Bruning, C.A., Acker, C.I., Neto, J.S. and Nogueira, C.W. (2013). 2-Phenylethynylbutyltellurium enhances learning and memory impaired by scopolamine in mice. *Behav Pharmacol*, 24:249–254.
- Sultana, Z., Takaoka, J. and Koga, T. (2013). Resource value differentially affects fighting success between reproductive and non-reproductive seasons, *J Ethol.*, 31:203–209.
- Susswein, A. J., Katzoff, A., Miller, N. and Hurwitz, I. (2004). Nitric oxide and memory. *Neuroscientist*. 10(2):153-62.
- Tapsell. (2006). Health benefits to herbs and spices: the past, the present and future. *Medicinal Journal of Austria* 1:170-190
- Togo, T., Katsuse, O. and Iseki, E. (2004). Nitric oxide pathways in Alzheimer's disease and other neurodegenerative dementias. *Neurol Res*, 26: 563–566.
- Tota, S, Nath, C, Najmi, A.K., Shukla, R. and Hanif, K. (2012a.) Inhibition of central angiotensin converting enzyme ameliorates scopolamine induced memory impairment in mice: role of cholinergic neurotransmission, cerebral blood flow and brain energy metabolism. *Behav Brain Res.*, 232:66-76.
- Varshney, R. and Kale, R.K. (1990). Effects of Calmodulin Antagonists on Radiation-induced Lipid Peroxidation in Microsomes. *Int'l j Rad. Biol.*, 58:733-743.
- Walton, N. M., Shin, K., Tajinda, C. L., Heusner, J. H., Kogan, S., Miyake, C. Q., Tamura K 2012. Adult neurogenesis transiently generates oxidative stress. *PLoS One*, 7:e35264
- Wang, W., Li, S., Dong, H., Lv, S. and Tang, Y. (2009). Differential impairment of spatial and non-spatial cognition in a mouse model of brain aging. *Life Sci.*, 85: 127–135.
- Wimo, A., Jönsson, L., Bond, J., Prince, M. and Winblad, B. (2013). The worldwide economic impact of dementia 2010. *Alzheimer disease international. Alzheimers Dement*, 9 (1):1–11.
- Xingxuan H., Huang, Y., Li, B., Gong, C., and Schuchman, E. H. (2010). Deregulation of sphingolipid metabolism in Alzheimer's disease. *Neurobiology of Aging* 31: 398–408.
- Zaki, N., Efimov, D. and Berengueres, J. (2013). Protein complex detection using interaction reliability assessment and weighted clustering coefficient. *BMC Bioinformatics* 14:163.